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**(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)**

## **AHD (Aminohydantoin Hydrochloride) ELISA Kit**

Catalog No: E-FS-E004

96T

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Phone: 240-252-7368(USA) 240-252-7376(USA)

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## Test principle

This kit uses Indirect-Competitive-ELISA as the method. It can detect Aminohydantoin hydrochloride (AHD) in samples, such as honey, fish, shrimp, birds, liver and other samples. This kit is composed of Micro ELISA Plate, HRP conjugate, antibody, standard and other supplementary Solutions. The micro-plate provided in this kit has been pre-coated with AHD. During the reaction, AHD in the samples or standard competes with AHD on the solid phase supporter for sites of AHD antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each micro plate well, and TMB substrate is for color development. There is a negative correlation between the OD value of samples and the concentration of AHD. The concentration of AHD in the samples can be calculated by comparing the OD of the samples to the standard curve.

## Technical indicator

**Sensitivity:** 0.02 ppb (ng/mL)

**Reaction mode:** 25°C, 45min~15min

**Detection limit:** Tissue/liver---0.04ppb, Honey/milk/casing---0.04ppb,  
Milk powder/egg/egg powder---0.04ppb

As fish/shrimp tissue are disturbed by some factor, detection limit is 0.06ppb

**Cross-reactivity:** AHD---100%, AOZ/SEM/AMOZ<0.1%,

**Sample recovery rate:** Tissue/liver---85% ± 25%, Honey/milk/casing---80% ± 20%  
Milk powder/ egg/egg powder---85% ± 25%

## Kits components

Item	Specifications
ELISA Micro-plate	96 wells
Standard Liquid	1mL each (0ppb,0.02ppb,0.03ppb,0.18ppb,0.54ppb,1.62ppb)
High Concentrated Standard (100ppb)	1mL
DerivatizationSolution(black cap)	10mL
HRPConjugate(red cap)	5.5mL
Antibody Working Solution(blue cap)	5.5mL
Substrate Solution A(white cap)	6mL
Substrate Solution B(black cap)	6mL
Stop Solution(yellow cap )	6mL
20×Concentrated Wash Buffer(white cap)	40 mL
2×RedissolveBuffer(yellow cap )	50mL
Product Description	1 copy

## Other supplies required

**Instruments:** Micro-plate reader, Printer, Homogenizer, Oscillators, Centrifuge, Graduated pipette, Balance (sensitivity 0.01g).

**High-precision transfer pipette:** single channel (20-200 $\mu$ L, 100-1000 $\mu$ L), Multichannel (300 $\mu$ L).

**Reagents:** Ethyl acetate, N-hexane, Sodium hydroxide, Concentrated hydrochloric acid,  $K_2HPO_4 \cdot 3H_2O$ , Potassium nitroprusside ( $K_2Fe(CN)_5(NO) \cdot 2H_2O$ ), Glucuronidase, Zinc sulfate ( $ZnSO_4 \cdot 7H_2O$ ).

## Experimental preparation

Bring all Solutions and samples to room temperature before use.

Open the microplate reader in advance (30min), preheat the instrument, and set the testing parameters.

**1. Sample pretreatment Notice:** experimental apparatus should be clean, and the pipette should be disposable to avoid cross-contamination during the experiment.

### 2. Solution preparation

Solution 1: 0.36M Potassium nitroprusside solution (milk, milk powder)

Dissolve 11.9g of Potassium nitroprusside to 100mL with deionized water

Solution 2: 1.04M Zinc sulfate solution (milk, milk powder)

Dissolve 29.8g of Zinc sulfate to 100mL with deionized water

Solution 3: 0.1M  $K_2HPO_4$

Dissolve 11.4g of  $K_2HPO_4 \cdot 3H_2O$  to 500mL with deionized water

Solution 4: 1M HCl

Dilute 8.6mL of Concentrated hydrochloric acid to 100mL with deionized water.

Solution 5: 1M NaOH

Dissolve 4g Sodium hydroxide to 100mL with deionized water

Solution 6: Re-dissolve solution (If sample is aquiform, no need dilute)

Dissolve 2 $\times$  Re-dissolve solution to 1 $\times$  Re-dissolve solution with deionized water for sample re-dissolution. Store at 4 $^{\circ}$ C, valid for 1 month.

### 3. Sample pretreatment procedure

#### 3.1 Pretreatment of milk (liquid):

- (1) Take 5mL milk into 50mL EP tube, add 250 $\mu$ L of Potassium nitroprusside solution (Solution 1) and oscillate for 30s, then add 250 $\mu$ L of Zinc sulfate solution (Solution 2) and oscillate for 30s, at last centrifuge at 4000r/min for 10min at 15 $^{\circ}$ C;
- (2) Removed 1.1mL of the supernatant to another centrifuge tube, add 4mL deionized water, 0.5mL of HCl (1M) and 100 $\mu$ L of derivatization Solution, oscillate for 3min;
- (3) Incubate at 37 $^{\circ}$ C (about 16h) for overnight, or water-bath at 50 $^{\circ}$ C for 3h;
- (4) Add 5mL of  $K_2HPO_4$  (0.1M), 0.4mL of NaOH (1M) and 5mL ethyl acetate, then oscillate for 5min;
- (5) Centrifuge at 4000r/min for 10min at room temperature;
- (6) Removed 2.5mL of supernatant to another centrifuge tube, dry with Nitrogen Evaporators at

50-60°C;

- (7) Dissolved the residue with 1 mL of n-hexane, add 1 mL of 1× Re-dissolve solution and mix fully by shaking for 30 s;
- (8) Centrifuge at 4000 r/min for 5 min at room temperature, take 50 μL for detection and analysis.

**Note: Sample dilution factor: 2, minimum detection dose: 0.04 ppb**

### 3.2 Pretreatment of milk and egg powder:

- (1) Weigh  $1 \pm 0.05$  g of homogenate milk into EP tube, add 4 mL deionized water, 0.5 mL HCl (1M) and 100 μL of derivatization solution, oscillate for 5 min;
- (2) Incubate at 37°C (about 16 h) for overnight, or water-bath at 50°C for 3 h;
- (3) Add 250 μL Potassium nitroprusside solution (Solution 1) and oscillate for 30 s, then add 250 μL of Zinc sulfate solution (Solution 2) and oscillate for 30 s, at last centrifuge at 4000 r/min for 10 min at 15°C;
- (4) Removed all the supernatant to another centrifuge tube, add 5 mL of K<sub>2</sub>HPO<sub>4</sub> (0.1M), 0.4 mL of NaOH (1M) and 5 mL of ethyl acetate, then oscillate for 5 min;
- (5) Centrifuge at 4000 r/min for 10 min at room temperature;
- (6) Removed 2.5 mL of supernatant to another centrifuge tube, dry with Nitrogen Evaporators at 50-60°C;
- (7) Dissolved the residue with 1 mL of n-hexane, add 1 mL of 1× Re-dissolve solution and mix fully by shaking for 30 s;
- (8) Centrifuge at 4000 r/min for 5 min at room temperature, take 50 μL for detection and analysis.

**Note: Sample dilution factor: 2, minimum detection dose: 0.04 ppb**

### 3.3 Pretreatment of honey, tissue, liver, casing, feed and egg:

- (1) Weigh  $1 \pm 0.05$  g of homogenate milk into EP tube, add 4 mL deionized water, 0.5 mL of HCl (1M) and 100 μL of derivatization Solution, oscillate for 5 min;
- (2) Incubate at 37 °C (about 16 h) for overnight, or water-bath at 50 °C for 3 h;
- (4) Remove all the supernatant to another centrifuge tube, add 5 mL of K<sub>2</sub>HPO<sub>4</sub> (0.1M), 0.4 mL of NaOH (1M) and 5 mL of ethyl acetate, then oscillate for 5 min;
- (3) Centrifuge at 4000 r/min for 10 min at room temperature;
- (4) Remove 2.5 mL of supernatant to another centrifuge tube, dry with Nitrogen Evaporators at 50-60°C;
- (5) Dissolve the residue with 1 mL of n-hexane, add 1 mL of 1× Re-dissolve solution and mix fully by shaking for 30 s;
- (6) Centrifuge at 4000 r/min for 5 min at room temperature, take 50 μL for detection and analysis.

**Note: Sample dilution factor: 2, minimum detection dose: 0.04 ppb**

### **Assay procedure**

Centrifuge the sample again after thawing before the assay. Bring all reagents to room temperature for 30 min before use. **All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming.** Dilute 40mL of concentrated wash buffer into 800mL wash working buffer with deionized or distilled water.

- 1. Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells.
- 2. Add sample:** add 50 $\mu$ L of Standard or Sample per well, then add 50 $\mu$ L of HRP conjugate to each well, then add 50 $\mu$ L of antibody working solution, cover the plate with sealer we provided, oscillate for 5s gently to mix thoroughly, incubate for 45min at 25 $^{\circ}$ C.
- 3. Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 250 $\mu$ L of wash working buffer to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
- 4. HRP conjugate:** add 100 $\mu$ L HRP conjugate to each well, shading light incubation for 30min at 25 $^{\circ}$ C.
- 5. Wash:** repeat step 3
- 6. Color Development:** add 50 $\mu$ L of substrate solution A to each well, and then add 50 $\mu$ L of substrate solution B. Gently oscillate for 5s to mix thoroughly. Incubate shading light for 15min at 25 $^{\circ}$ C.
- 7. Stop reaction:** add 50 $\mu$ L of stop solution to each well, gently oscillate for 5s.
- 8. OD Measurement:** determine the optical density (OD value) of each well at 450 nm with a micro-plate reader (the 450/630 nm double wavelength is recommended). This step should be finished in 10min after stop reaction.

### **Result analysis:**

**1. Absorbance (%) =  $A/A_0 \times 100\%$**

A: Average absorbance of standard or sample

$A_0$ : Average absorbance of 0 ppb Standard

**2. Drawing and calculation of standard curve**

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.

For this kit, it is more convenient to use professional analysis software for accurate and fast analysis on a large number of samples.

### **Notes**

1. Overall OD value will be lower when Solutions is not brought to room temperature before use or room temperature is below 25°C.
2. During the washing procedure, if the wells turn dry, it will lead to bad linear standard curve and poor repeatability, move on to the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. Micro ELISA plate should be covered by plate sealer. Avoid the Solutions to strong light.
5. Do not use expired kit and reagents of different batches.
6. TMB should be abandoned if it turns color. When OD value of standard (concentration: 0)  $< 0.5$  unit ( $A_{450nm} < 0.5$ ), it indicates Solution is deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.

### **Storage and valid period**

**Storage:** Store at 2-8°C. Avoid freeze / thaw cycles.

**Valid Period:** 1 year, production date is on the packing box.